

REVIEW

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REVIEW

Exploration of endophytic fungi for applications in agriculture and medicine

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Endophytic fungi are microbes that colonize inner healthy plant tissues without causing any disease symptoms on its host. They have been isolated from every plant species investigated so far. The functional roles of endophytes are manifold. They are known to confer considerable benefits to the host by producing substances that stimulate plant growth, enhanced resistance to biotic and abiotic stresses. Besides they produce secondary metabolites that inhibit both phytopathogens and clinically significant human pathogens. Till recently, 5000+ bioactive metabolites have been isolated from endophytic microbes. Therefore, endophytes are recognized as repository of bioactive metabolites and have been commonly term as "chemical synthesizers" inside the host. Many scientists believe that plants growing in lush tropical rainforests, where competition for light and nutrients is severe, are most likely to host the greatest number of bioactive endophytes than temperate parts of the worlds. Thus, considering the myriad of medicinal plants particularly in biodiversity rich regions, study of endophytic microbes associated with these plant resources might lead to the discovery of new and effective molecules with wide therapeutic applications. In the recent years endophytic microbes have also been proved beneficial in agriculture. Many endophytic microbes have been reported to produce growth hormones, secondary metabolites that inhibit plant pathogens, mitigate biotic and abiotic stresses. Therefore they have been found suitable to apply as bio-inoculants to promote plant growth and health. This review will therefore focus on importance of endophytic microbes and their applications in the field of agriculture and medicine.

Key words: Endophytes, diversity, antimicrobial metabolites, plant growth promotion

INTRODUCTION

As the global population continues to grow the demand for food and other agricultural products will be on an increasing trend. This is more challenging considering the drastic climate change which has led to the compounds issues like loss of soil fertility, pest and pathogens damage, abiotic stress and others.

Further, the emergence of new diseases, multi-drug resistance pathogenic microbes, needs new and therapeutic agents. Thus, research is necessary to solve these ever growing human problems in an innovative and comprehensive

manner. Endophytes are microorganisms that colonize inner healthy tissues and establish for all or part of their life cycle without causing any disease symptoms. Endophytes are quite common and found almost in every vascular plant (Rodriguez *et al.* 2009). Although various microbes exist as endophytes but the most commonly studied are the fungi. The frequency and diversity of fungal endophytes present in the host plant might vary based on the location and climatic condition of the area where the host is growing. Study of endophytic microflora had gained tremendous attention because they are the sources of many industrially, agriculturally and medicinally important compounds. Many endophytic microorganisms can produce the secondary metabolites which are similar to the one produced by the plant and can be used as alternate source of novel plant biomolecules which might help in the sustainable utilization and conservation of some of the

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economically important rare and endangered plants. For example Taxol, an anticancer agent obtained from leaves and bark of *Taxus brevifolia*, is a commercially important plant. The fungus *Taxomyces andreanae*, an endophyte isolated from *T. brevifolia*, have been found to produce taxol. Such findings attracted the scientific community of the world towards exploration of endophytes for various bioactive secondary metabolites.

The functional roles of endophytes are manifold. They are known to confer considerable benefits to the host by producing substances that stimulate plant growth, enhanced resistance to biotic and abiotic stress. Besides they produce secondary metabolites that inhibit both phytopathogens and clinically significant human pathogens. Till recently, 5000+ bioactive metabolites have been isolated from endophytic microbes. Therefore, endophytes are recognized as repository of bioactive metabolites and have been commonly term as “chemical synthesizers” inside the host. Many scientists believe that plants growing in lush tropical rainforests, where competition for light and nutrients is severe, are most likely to host the greatest number of bioactive endophytes than temperate parts of the worlds. Thus, considering the myriad of plants species particularly in biodiversity rich regions, study of endophytic microbes associated with these plant resources might lead to the discovery of new and effective molecules with wide therapeutic applications. In the recent years endophytic microbes have also been proved beneficial in agriculture. Many endophytic microbes have been reported to produce growth hormones, secondary metabolites that inhibit plant pathogens, mitigate biotic and abiotic stresses. Therefore they have been found application as bio-inoculants to promote plant growth and health. Further, different enzymes produced by endophytic fungi residing in different plant hosts plays many useful roles like enhanced resistance to different biotic and abiotic stresses by producing various bioactive compounds such as alkaloids, terpenes, flavonoids etc. (Firakova *et al.* 2007). Lignocellulolytic enzymes are one of the most important enzymes produced by various groups of fungi, which can degrade lignocellulose compounds. Recently, some endophytic fungi have also been reported to exhibit lignocellulolytic enzymes (Toghueo *et al.* 2017;Kaul *et al.* 2014). Lignocellulose is a collective name of three polymers, cellulose (35-50%), hemicellulose (20-

35%) and lignin (10-25%) (Isikgor *et al.* 2015). Cellulase is an industrially important enzyme which can degrade cellulose by hydrolysing α -1,4-glycosidic bonds. Degradation of lignocellulose compounds is very significant because they can be converted to agriculturally important compost. Therefore, this review will focus on diversity of endophytic fungi in various plant species and their applications in the field of agriculture and medicine. A brief highlight on research outcome of endophytic fungi has been projected and discussed.

Diversity of endophytic fungi across plant species

Endophytic fungi are known to be ubiquitous and found to be associated with almost all the plant species of the planet. They have been isolated from all vascular plant investigated so far (Sturzand Nowak, 2000; Arnold *et al.* 2000). However, tropical and subtropical plants have been reported to have higher endophytic fungal assemblages (Suryanarayanan and Rajagopal, 2000). This might be due to favorable climatic conditions such as increased rate of precipitation, humidity and nutrient availability in tropics. Tropical rainforest are one of the most biologically diverse terrestrial ecosystems on the planet. This region harbors the maximum plant species in the world. However, very few of them have been studied for endophytic fungi. Thus, there exist great opportunity to explore those cryptic fungi which might contribute substantially to global fungal diversity. Endophytic fungal diversity has been studied from various healthy tissues of the host such as leaves, petioles, barks, stems, roots and seeds. Some common plant species investigated for endophytic fungi are *Azadirachta indica*, *Aegle marmelos*, *Holarrhena antidysenterica*, *Terminalia arjuna*, *Ocimum sanctum*, *Vitex negundo*, *Emblica officinalis*, *Coffea robusta* etc. Several fungal species have been reported as endophytes. Many of them closely resemble plant pathogenic fungi. Some of the common fungi isolated as endophytes are *Acremonium* sp., *Alternaria alternata*, *Arthrinum* sp., *Aspergillus niger*, *Aspergillus fumigatus*, *Chaetomium globosum*, *Cladosporium* sp., *Colletotrichum* sp., *Curvularia* sp., *Diaporthe* sp., *Fusarium* spp., *Muscodora albus*, *Nigospora* sp., *Penicillium* sp., *Pestalotiopsis* sp., *Phomopsis* sp., *Phyllosticta*, *Rhizoctonia* sp., *Trichoderma* sp., *Xylaria* sp., and many more. Some plant species

along with their associated endophytic fungi is presented in Table 1.

We also studied diversity of endophytic fungi from various plant species. *Taxus baccata* a well known anticancer plant was investigated for endophytic fungi. Bark of *T. baccata* collected from Arunachal Pradesh, India revealed rich diversity of endophytic fungi. The dominant endophytic fungi isolated from the collected samples were *Alternaria alternata*, *Chaetomium globosum*, *Fusarium moniliforme*, *Gliocladium* sp., *Nigrospora oryzae*, *Penicillium* sp., *Trichoderma viridae* and sterile mycelia (Tayung and Jha, 2010). Endophytic fungi were also studied from three ethno-medicinal plants namely *Andrographis paniculata*, *Acorus calamus*, and *Drynaria quercifolia* collected from Similipal Biosphere Reserve, India. The isolated endophytic fungal genera were *Alternaria*, *Aspergillus*, *Curvularia*, *Fusarium* and *Penicillium* (Mohanta *et al.* 2008). Endophytic fungi were studied from an invasive plant, *Ipomea carnea*. A total of 69 isolates belonging to ten taxa comprising 1.45% Zygomycetes, 10.14% Coelomycetes, 62.32% Hypomycetes, 18.84% sterile mycelia and 7.25% unidentified species were obtained from leaves, stems and seeds. Species of *Curvularia*, *Aspergillus*, *Fusarium*, *Colletotrichum* and sterile fungus were isolated as dominant endo-phytes. Colonization frequency of *Curvularia* (7.25%) was highest which was isolated from all the tissues. Furthermore, endophytic fungi were also studied from two ethnomedicinal plants namely *Solanum rubrum* and *Morinda pubescence*. A total of 458 endophytic isolates were obtained from leaves, stems and fruit tissues. The dominant endophytic fungi belong to genera *Aspergillus*, *Colletotrichum*, *Curvularia* and Mycelia sterilia (Jena and Tayung, 2013). Recently endophytic fungi from three medicinal plants of NE India were investigated. Altogether, 56 endophytic fungal isolates were obtained from surface-sterilized leaf fragments of *Houttuynia cordata*. The endophytes consisted of fungi belonging to genera *Colletotrichum*, *Curvularia*, *Bipolaris*, *Corynespora*, and *Pseudozyma* and non-sporulating fungi categorized as mycelia sterilia. (Talukdar *et al.*, 2020). In another study, a total of 84 endophytic fungal isolates were isolated from 150 segments of leaf tissues of *Eyngium foetidum* (Fig.1). Dominant endophytes were found to be fungi belonging to the genus *Colletotrichum*, followed by non-sporulating members grouped under mycelia

sterilia. Other fungal genera that were isolated as endophytes were *Scopulariopsis*, *Cladosporium*, *Stemphylium*, *Penicillium* and *Alternaria*. Again from healthy leaves tissue of *Zanthoxylum oxyphyllum* a medicinal plant widely been used by the local tribal communities of Assam, *Colletotrichum* species was isolated as dominant endophytes along with other fungal genera like *Fusarium*, *Curvularia*, *Aspergillus*, *Corynespora* and isolates belonging non-sporulating fungi categorised as Mycelia Sterilia (Talukdar and Tayung, 2021). Besides, endophytic fungi were also isolated from orchid tissues, seeds of *Cassia alata*, *Oryza sativa* seeds and lichen thalli (Padhi and Tayung, 2015; Roy *et al.*, 2021). The list of plant species investigated for endophytic fungi along with its associated endophytes is presented in Table 2. Tropical and sub-tropical rainforest harbor rich diversity of plant species. Owing to its favorable climatic conditions these plants serves as the reservoir for obtaining novel endophytic microbes. Diminishing of these plant resources will also eliminate the associate endophytes. Hence research priority should be directed to study them before they are exploited or get extinct.

Antimicrobial metabolites from endophytic fungi

The developed world has seen an explosion in the incidence and reporting of multi-resistant drug infections by pathogenic microorganisms and emerging of new diseases. This has necessitated the needs for new and effective antimicrobial agents to control human health problems. Without new antibiotics, medicine will change beyond recognition as most of the currently used antibiotics are getting resistant. The failure to bring new antimicrobial agents to the market partly reflects a failure of discovery. This is because many of the reputed pharma companies have stop finding for antibiotics research and have diverted their attention to invest medicines for diseases like diabetes, anti-aging and heart. As a result, the search for new antibiotics is almost shrinking gradually. Microorganisms have been good sources of antibiotics since decades. Among microbes, endophytic fungi are known for their production of diverse biologically active metabolites. Many endophytic fungi produce antimicrobial metabolites in culture that are active against human and plant pathogens.

In recent years, endophytic fungi colonizing medicinal plants are of special interest because of

the fact that such microbes have produced similar metabolites as their respective host, sometime displaying more biological activity. There is a need to investigate fungal endophytes from medicinal plants because it has been hypothesized that these plants harbor some distinct and rare microbes that mimic the chemistry of their respective hosts and synthesize identical bioactive natural products or derivatives that are more bioactive than the one produced by the host. Strobel and Daisy (2003) have necessitated the need to study plants growing in unique environmental settings having ethno medicinal uses, extreme age or interesting endemic locations because they are expected to harbor novel endophytes that may produce unique metabolites having diversified applications. Wang *et al.* (2007) have demonstrated that the endophytes isolated from these plants are excellent producer of strong fungicidal and bactericidal metabolites. The production of antimicrobial metabolites from endophytic fungi around the world has been well document by Strobel and Daisy (2003). Endophytic *Phoma* sp., isolated from different medicinal plants has been reported to be a promising source of antimicrobial compounds. A new alpha-tetralone derivative -3,6,7-trihydroxy-alpha-tetralone together with cercosporamide, beta-sitosterol and trichodermin was reported to be produced by *Phoma* sp. endophytic in *Arisaema erubescens*. These isolated compounds exhibited antifungal and antibacterial activity against pathogenic fungi *Fusarium oxysporum*, *Rhizoctonia solani*, *Colletotrichum gloeosporioides* and *Magnaporthe oryzae* as well as against two plant pathogenic bacteria *Xanthomonas campestris* and *Xanthomonas oryzae* (Wang *et al.* 2012). Similarly, *Phoma* sp., endophyticin *Saurauia scaberrinae* is known to produce Phomodione, an usnicacid derivative. Phomodione was found to be effective at a minimum inhibitory concentration of 1.6 mg/ml against *Staphylococcus aureus* (Hoffman and Arnold, 2008). A terpenoid compound with known antibacterial activity was obtained from the ethyl acetate fraction of *Phomopsis* sp. an endophyte of *Plumeria acutifolia* Poiret plant (Nithya and Muthumary, 2011). Ethyl acetate extract of *Xylaria* sp. isolated from *Piper aduncum* also produced two new presilphiperfolane sesquiterpenes with antifungal activity (Silva *et al.* 2010). Liu *et al.* (2008) reported *Xylaria* sp. YX-28 from *Ginkgo biloba* producing 7-amino-4-methylcoumarin which showed antibacterial and antifungal activity against

many pathogenic organisms. Another endophytic *Fusarium* sp. produced novel antifungal antibiotic Fusarielin A and three related compounds. The mangrove endophytic fungus, *Phomopsis* sp. ZSUH76 produced three new metabolites *Phomopsin* A, B, C and known cytosporone B and cytosporone C. The latter possessed antifungal activity against *Candida albicans* and *F. oxysporum*. A few works on antimicrobial metabolites have been undertaken from India. Raviraja *et al.* (2006) have evaluated antimicrobial activity of some endophytic fungi isolated from medicinal plants of the Western Ghats of India. Elavarasi *et al.* (2012) have reported endophytic fungus, *Fusarium* sp. from mangrove producing volatile antimicrobial metabolites 1-Dodecanol, 2-methyl. Similar work on antimicrobial metabolites from endophytic fungi isolated from various plant species were also reported by us (Mohanta *et al.* 2008; Tayung and Jha, 2010; Tayung *et al.* 2012, Padhiand Tayong, 2013). Ethyl acetate extract obtained from an endophytic fungus, *Fusarium solani* isolated from bark of *Taxus baccata* showed both antibacterial and antifungal activity. The metabolites were identified as 1-tetradecene, 8-octadecanone, 8-pentadecanone, octylcyclohexane and 10-nonadecanone (Tayung *et al.* 2011). Recently, antimicrobial and anti-quorum sensing metabolites identified as tyrosol was isolated from an endophytic fungus, *Colletotrichum coccodes* obtained from healthy leaf tissues of *Houttuynia cordata* (Talukdar *et al.* 2021). Similarly, endophytic fungi isolated from *Zanthoxy-lumoxyphyllum* a medicinal plant showed promising antimicrobial activity against selected clinically significant human pathogenic test organisms. (Fig.2) (Talukdar and Tayung, 2021). Such finding suggested that extensive and scientific investigation of medicinal plants for endophytic microbes might leads to the discovery of potent endophytic strains with antimicrobial activity that could be developed into new and effective antimicrobial agents.

Antimicrobial metabolites from endolichenic fungi

Endolichenic fungi are organisms that live in close association with the alga in the thalli of lichen are analogous to that of plant endophytes inhabiting the intercellular spaces of the hosts. Similar to fungal endophytes, all lichen species studied so far have been found to be colonized by endolichenic fungi (Arnold *et al.* 2009; Tripathi and

Joshi 2015; Suryanarayanan and Thirunavukkarasu, 2017). However, considering the myriad of lichen species in the planet, only a tiny fraction of them have been studied for endolichenic fungi. Studies carried out by Bates *et al.* (2012) showed that numerous fungi are present in lichen thalli. Additionally, like the endophytic fungi, they are reported to produce an array of secondary metabolites such as alkaloids, quinones, furanones, pyrones, benzopyranoids, xanthenes, terpenes, steroids, peptides and allylic compounds (Paranagama *et al.* 2007; Yang *et al.* 2012; Li *et al.* 2015; Kellogg and Raja 2017). The metabolites obtained from endolichenic fungi have been demonstrated to produce various antibiotics and natural bioactive compounds with multiple applications (Wu *et al.*, 2011). However, relatively very few studies have been undertaken on endolichenic fungi, and the substances they produced have not been investigated in detail for their bioactivity and therapeutic potentials.

In recent years several bioactive metabolites have been reported from endolichenic fungi. Some of these metabolites showed promising antibacterial, antifungal, cytotoxic and antioxidant activities (Kellogg and Raja, 2017; Suryanarayanan and Thirunavukkarasu, 2017). Endolichenic fungus identified as *Aspergillus niger* isolated from lichen thallus of *Rocella montagnei* showed antibacterial activity against wide range of bacterial pathogens (Logesh *et al.* 2012). Amburic acid obtained from an endolichenic fungus, *Pestalotiopsis* sp. isolated from lichen thalli of *Clavarioides* sp., has shown antibacterial activity (Deng *et al.* 2009). Similarly, metabolites isolated from lichen thalli of *Usnea cavernosa* have showed antifungal activity against some clinically significant human pathogenic fungi (Paranagama *et al.* 2007). We also investigated several lichen species for endolichenic fungi and reported some promising metabolites with antimicrobial activity. Endolichenic fungi were studied from lichen thalli of *Parmelia* sp. A total of 19 distinct endolichenic fungi were obtained from surface sterilized fragments of *Parmelia* thalli. The dominant fungi belonged to genera *Phomopsis*, *Aspergillus*, *Penicillium* and *mycelia sterilia*. Some of the isolates showed significant antimicrobial activity against wide range of pathogenic microbes. We reported an undescribed substituted dihydroxanthene-1,9-dione, named funiculosone, obtained from the culture filtrates of *Talaromyces funiculosus*. The fungus was isolated as

endolichenic fungus from surface sterilized lichen fragments of *Diorygmahiero glypticum*. The metabolites showed promising antibacterial activity against *Escherichia coli* and *Staphylococcus aureus*. Funiculosone also showed anticandidal activity against *Candida albicans* with an IC₅₀ 35 µg/mL (Padhi *et al.* 2019). Furthermore, metabolites aspergillone isolated from culture filtrates of an endolichenic fungus *Aspergillus niger* obtained from lichen thallus *Parmotrem aravum* showed promising activity against a panel of human, plant, food borne and fish pathogens (Padhi *et al.* 2020). Such finding indicated that endolichenic fungi harboring lichen thalli may be a good source of new and effective antimicrobial metabolites.

Plant growth promotion by endophytic fungi

Endophytic fungi may directly or indirectly contribute plant growth and health. They have been known to increase the seed germination rate, seed elongation, and solubilize nutrients for uptake. Fungal endophytes also contribute significantly to plant growth by producing phyto-hormones, such as auxin and cytokinin, as well as in other products such as phosphatases that solubilized insoluble phosphorus and make it available to the plants (Tsavkelova *et al.* 2007). Many endophytic fungi have been reported to produce growth hormones like IAA and Gibberellin (Ismail *et al.* 2016) and also fix atmospheric nitrogen (Yang *et al.* 2015). Some endophytes have also been reported to solubilized phosphate and produce iron chelating metabolites called siderophores (Potshangbam *et al.* 2017). Soil-borne plant pathogens continue to be a major threat to agricultural development and productivity worldwide. Several plant disease control methods have been implemented to protect crops against a wide range of phyto-pathogens but most of the approaches tend to have one or the other side effect associated with them. Many endophytic fungi have been reported to produce secondary metabolites which are inhibitory to phyto-pathogenic microbes. Therefore the recent development of screening technologies has revealed the great potential of fungal endophytes for producing novel biologically active compounds with promising medicinal or agricultural applications (Zhang *et al.* 2006; Aly *et al.* 2011; Wu *et al.* 2015).

There are many examples of secondary metabolites from endophytic fungi which are being used, or have promise to be used, against

Table 1: Endophytic fungi isolated from some selected plants species

Host plant	Isolated endophytic fungi	Reference
<i>Azadirachta indica</i>	<i>Acremonium</i> sp.	Tejesvi <i>et al.</i> , 2006
<i>Aegle marmelos</i>	<i>Alternaria alternate</i> <i>Aspergillus niger</i>	Gond <i>et al.</i> , 2007
<i>Butea monosperma</i>	<i>Acremonium strictum</i> <i>Asteromella andrewsii</i>	Tejesvi <i>et al.</i> , 2006
<i>Cajanus cajan</i>	<i>Fusarium solani</i> , <i>Fusarium oxysporum</i> , and <i>Fusarium proliferatum</i>	Zhao <i>et al.</i> 2012
<i>Capsicum annum</i>	<i>Alternaria alternata</i>	Devari <i>et al.</i> 2014
<i>Coffea arabica</i>	<i>Aspergillus versicolor</i>	Sette <i>et al.</i> , 2006
<i>Clerodendrum serratum</i>	<i>Aspergillus</i> sp.	Banerjee <i>et al.</i> , 2009
<i>Cinchona ledgeriana</i>	<i>Penicillium</i> , <i>Fomitopsis</i> and <i>Arthrinium</i>	Maehara <i>et al.</i> 2013 and Maehara <i>et al.</i> 2011
<i>Catharantus roseus</i>	<i>Fusarium oxysporum</i> , <i>Talaromyces radicus</i> , and <i>Eutypella</i> spp	Palem <i>et al.</i> , 2016
<i>Centella asiatica</i>	<i>Colletotrichum</i> sp. <i>Glomerella</i> sp.	Rakotoniriana <i>et al.</i> , 2008
<i>Embelia ribes</i>	<i>Aspergillus</i> sp. <i>Bipolaris spicifera</i> <i>Fusarium</i> sp. <i>Pestalotia</i> sp	Mokaya and Peter., 2016
<i>Fritillaria cirrhosa</i>	<i>Fusarium redolens</i>	Pan <i>et al.</i> 2015
<i>Gerbera piloselloides</i>	<i>Colletotrichum</i> sp. <i>Alternaria</i> sp. <i>Penicillium</i> sp. <i>Fusarium</i> sp.	Caruso <i>et al.</i> , 2020
<i>Ginkgo biloba</i>	<i>Fusarium oxysporum</i>	Cui <i>et al.</i> 2012
<i>Houttuynia cordata</i>	<i>Colletotrichum</i> sp., <i>Fusarium</i> sp. <i>Lasiodiplodia pseudotheobromae</i> <i>Xylariales</i> sp.	Aramsirujwet & Kitpreechavanich, 2016
<i>Musa acuminata</i>	<i>Colletotrichum gloeosporioides</i> C. Muase, <i>Guignardia cocoicola</i>	Wipornpanet <i>et al.</i> , 2001
<i>Leucas aspera</i>	<i>Colletotrichum</i> sp.	Banerjee <i>et al.</i> , 2009
<i>Macleaya cordata</i>	<i>Fusarium proliferatum</i>	Wang <i>et al.</i> 2014
<i>Passiflora incarnata</i>	<i>Alternaria alternata</i> , <i>Colletotrichum capsici</i> , and <i>Chryseobacterium taiwanense</i>	Seetharaman <i>et al.</i> , 2017
<i>Solanum nigrum</i>	<i>Fusarium</i> sp. <i>Alternaria</i> sp.	Khan <i>et al.</i> , 2015
<i>Terminalia arjuna</i>	<i>Cladosporium</i> sp. <i>Chloridium</i> sp.	Tejesvi <i>et al.</i> , 2006

(contd. part table 1)

<i>Ocimum sanctum</i>	<i>Gliocladiumdelequescens</i> <i>Bacillus subtilis</i> <i>Aspergillus niger</i> <i>Alternaria alternata</i> <i>Bipolaris maydis</i> <i>Fusarium verticillioides</i>	Chowdhary and Nutan, 2015
<i>Rauvolfia serpentine</i>	<i>Colletotricumgloeosporioides</i> , <i>Penicillium sp.</i> , <i>Aspergillus awamori</i>	Nath <i>et al.</i> , 2015
<i>Vitex negundo</i>	<i>Fusarium sp.</i> <i>Papulaspora sp.</i>	Banerjee <i>et al.</i> , 2009 Raviraja <i>et al.</i> , 2006

Table 2: List of some selected plant species studied for endophytic fungi

Plant name	Family	Habit	Economic importance
<i>Houttuynia cordata</i>	Saururaceae	herb	Medicinal
<i>Eryngium foetidum</i>	Apiaceae	herb	Medicinal
<i>Zanthoxylum oxyphyllum</i>	Rutaceae	shrub	Essential oil, medicinal
<i>Solanum rubrum</i>	Solanaceae	shrub	Medicinal
<i>Morinda pubescens</i>	Rubiaceae	herb	Medicinal
<i>Taxus baccata</i>	Taxaceae	tree	Anticancer
<i>Ipomoea carnea</i>	Convolvulaceae	shrub	Medicinal/invasive
<i>Rhynchosstylis retusa</i>	Orchidaceae	epiphytic	Medicinal
<i>Shorea robusta</i>	Dipterocarpaceae	tree	Timber/medicinal
<i>Cassia alata</i>	Fabaceae	shrub	Medicinal/invasive
<i>Oryza sativa</i>	Poaceae	herb	Food
<i>Parmelia sp. (Lichen)</i>	Parmeliaceae	epiphytic	Medicinal
<i>Parmotrema ravum (Lichen)</i>	Parmeliaceae	epiphytic	Medicinal

pathogens and pests of horticultural and agricultural plants. Endophytic fungi have also been reported to improve the uptake of macronutrients, such as phosphorus, nitrogen, potassium and magnesium, or micronutrients, such as zinc, iron, and copper, from the soil and organic matter and increase the supply of these nutrients to the plant host (Rana *et al.* 2020). Garcia *et al.* (2015) reported that inoculation of endophytic fungal isolate, *Trichoderma asperellum* significantly reduced the use of phosphorus fertilisation in onion (*Allium cepa*). Similar results were obtained by Baron *et al.* (2018) where inoculated maize (*Zea*

mays) with *Aspergillus sydowii*, and the plants that interacted with the fungus accumulated significantly higher amounts of P in their tissues even when receiving lower fertilisation doses. Production of IAA and GA by endophytic fungi *Phoma glomerata* and *Penicillium sp.* isolated from cucumber plant (Waqau *et al.* 2012). Khan *et al.* (2008) reported gibberellin production by *Penicillium citrinum* IR-3-3, a fungus isolated as endophyte from dune plants. In our study also we have isolated many endophytic fungi from various plant species that showed plant growth promoting activities. In one the study endophytic fungi



Fig. 1 : Endophytic fungi growing out from the surface sterilized fragments in PDA medium(Source: Talukdar and Tayung, 2021)

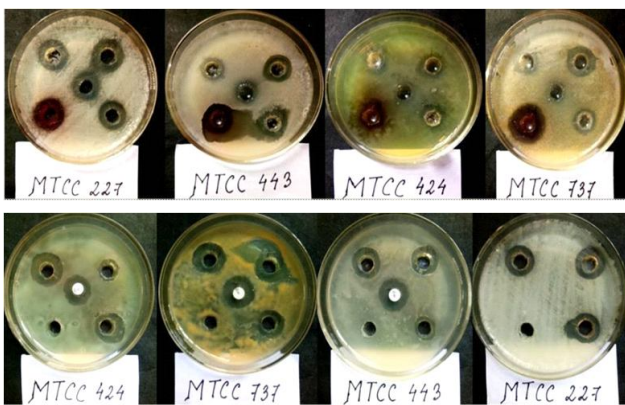


Fig. 2 : Antimicrobial activities of some endophytic fungi against some test pathogens (source: Talukdar and Tayung, 2021)

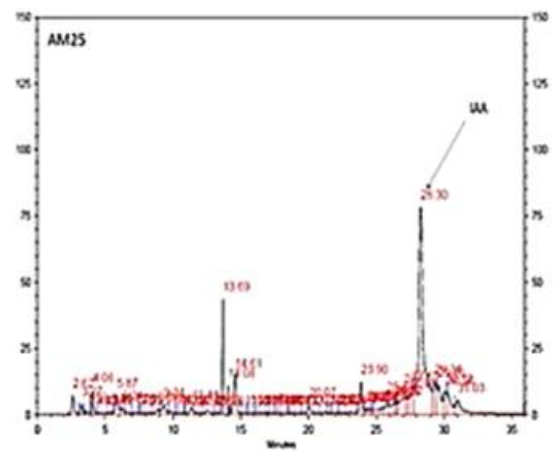
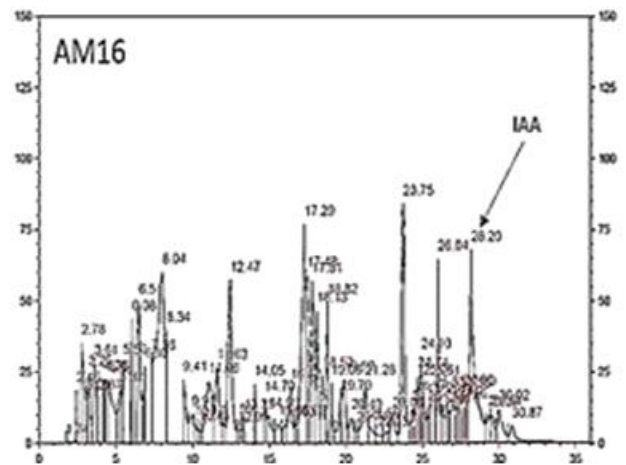


Fig. 4 : HPLC chromatogram showing presence of IAA in the crude metabolites obtained from endophytic fungi (Source: Roy et al., 2021)

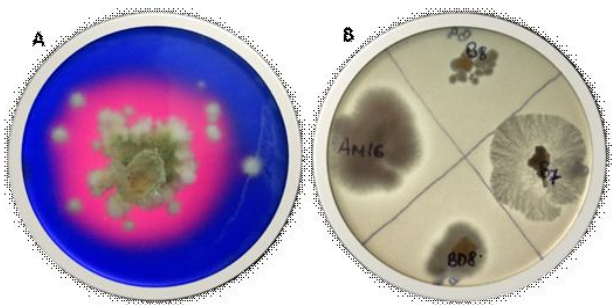


Fig. 3 : Siderophore (A) and phosphate solubilizing (B) activity of endophytic fungi isolated from seeds of indigenous rice cultivars (Source: Roy et al., 2021)

associated with seeds of some indigenous rice varieties of North East, India were determined for IAA activity *in-vitro* and antifungal activity against rice pathogen, *Magnaporthe grisea*. Among

several isolates, colonization frequency of *Fusarium* was found to be highest and was isolated from seeds of almost all the varieties. The isolates showed good IAA production in the medium amended with tryptophan and displayed antifungal activity against *Magnaporthe grisea* (Roy *et al.* 2021). Some of the fungal isolates also showed siderophore and phosphate solubilizing activity *in-vitro* (Figs. 3 & 4). Endophytic fungi isolated from seeds of *Cassia alata* were also investigated from plant growth promotion traits. High colonization of *Fusarium* species were observed in the seeds collected from different sampling sites. All of the isolates good showed IAA activity *in-vitro* (Data unpublished). Such results exemplify that endophytic fungi inhabiting plant species may be exploited for sustainable agriculture to improve plant growth and health and increase productivity.

Conclusions and future perspectives

At present the world is facing many problems and among them global food crisis, emerging of new diseases and antibiotic resistance pathogenic microbes are major concern. To solve these growing human problems in sustainable method, there is need to discover new therapeutic agents to combat drug resistant microbes and explore bioinoculants to be used in agriculture so as to improve plant growth and health. Endophytic fungi have been reported to produce numerous secondary metabolites with biological activities. The ability of endophytic fungi to produce metabolites with promising antimicrobial activity and plant growth promotion traits has proven to be boon for the present scientific community to solve the ever growing human problems. As stated in the review endophytic fungi are beneficial because they provide new and effective bioactive metabolites for applications in agriculture and medicines. Considering the myriads plant species on the planet, exploration of endophytic fungi from different ecological niches may lead to the discovery of potent endophytes with multiple applications.

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REFERENCES

- Aly, A.H., Debbab, A., Proksch, P. 2011. Fungal endophytes: unique plant inhabitants with great promises. *Appl. Microbiol. Biotechnol.* **90**: 1829-1845.
- Aramsirujijwet, Y., Gumlangmak, C., Kitpreechavanich, V. 2016. Studies on antagonistic effect against plant pathogenic fungi from endophytic fungi isolated from *Hottuyenia cordata* Thunb and screening for Siderophore and indole-3-acetic acid production. *Asia-Pacific J. Sci. Technol.* **21**: 55-66.
- Arnold, A.E., Maynard, Z., Gilbert, G.S., Coley, P.D., Kursar, T.A. 2000. Are tropical fungal endophytes hyperdiverse? *Ecol.Lett.* **3**: 267-274.
- Arnold, A.E., Miadlikowska, J., Higgins, K.L., Sarvate, S.D., Gugger, P., Way, A., Lutzoni, F. 2009. A phylogenetic estimation of trophic transition networks for ascomycetous fungi: are lichens cradles of symbio-trophic fungal diversification? *Syst.Biol.* **58**: 283-297.
- Banerjee, D., Manna, S., Mahapatra, S., Pati, B. 2009. Fungal endophytes in three medicinal plants of Lamiaceae. *Acta Microbiologic.Immunologic. Hung.* **56**: 243-250.
- Baron, N.C., Costa, N.T.A., Mochi, D.A., Rigobelo, E.C. 2018. First report of *Aspergillus sydowii* and *Aspergillus brasiliensis* as phosphorus solubilizers in maize. *Annal.Microbiol.* **68**: 863-870.
- Bates, S.T., Donna, B.L., Lauber, C.L., Walters, W.A., Knight, R. and Fierer, N. 2012. A preliminary survey of lichen associated eukaryotes using pyrosequencing. *The Lichenologist* **44**: 137-146.
- Caruso, G., Abdelhamid, M. T., Kalisz, A., Sekara, A. 2020. Linking endophytic fungi to medicinal plants therapeutic activity. A case study on Asteraceae. *Agriculture* **10**: 286.
- Chowdhary, K., Kaushik, N. 2015. Fungal endophyte diversity and bioactivity in the Indian medicinal plant *Ocimum sanctum* Linn. *Plos one* **10**: e0141444.
- Cui, Y., Yi, D., Bai, X., Sun, B., Zhao, Y., Zhang, Y. 2012. Ginkgolide B produced endophytic fungus (*Fusarium oxysporum*) isolated from *Ginkgo biloba*. *Fitoterapia* **83**: 913-920.
- Deng, B.W., Liu, K.H., Chen, W.Q., Ding, X.W., Xie, X.C. 2009. *Fusarium solani*, Tax-3, a new endophytic taxol-producing fungus from *Taxus chinensis*. *World J. Microbiol.Biotechnol.* **25**: 139-143.
- Devari, S., Jaglan, S., Kumar, M., Deshidi, R., Guru, S., Bhushan, S., Shah, B. A. 2014. Capsaicin production by *Alternaria alternata*, an endophytic fungus from *Capsicum annuum*; LC-ESI-MS/MS analysis. *Phytochemistry* **98**: 183-189.
- Elavarasi, A., Rathna, G.S. and Kalaiselvam, M. 2012. Taxol producing mangrove endophytic fungi *Fusarium oxysporum* from *Rhizophora annamalayana*. *Asian Pac. J. Trop. Biomed.* **2**: S1081-S1085.
- Evans, M. N., Onderi, P. O. 2016. Isolation of endophytes and bioactive assays for *Embelia* species. *Eur. J. Biol. Med. Sci. Res.* **4**: 69-79.
- Firakova, S., Sturdikova, M., Muckova, M. 2007. Bioactive secondary metabolites produced by microorganisms associated with plants. *Biologia* **62**: 251-257.
- Garcia, N.F.L., Santos, F.R.D.S., Gonçalves, F.A., Paz, M.F.D., Fonseca, G.G., Leite, R.S.R. 2015. Production of α -glucosidase on solid-state fermentation by *Lichtheiariamosa* in agroindustrial residues: Characterization and catalytic properties of the enzymatic extract. *Electronic J. Biotechnol.* **18**: 314-319.
- Gond, S. K., Verma, V. C., Kumar, A., Kumar, V., Kharwar, R. N. 2007. Study of endophytic fungal community from different parts of *Aegle marmelos* Corraea (Rutaceae) from Varanasi (India). *World J. Microbiol. Biotechnol.* **23**: 1371-1375.
- Hoffman, M.T., Arnold, A.E. 2008. Geographic locality and host identity shape fungal endophyte communities in cupressaceous trees. *Mycologic.Res.* **112**: 331-344.

- Isikgor, F.H., Becer, C.R. 2015. Lignocellulosic biomass: a sustainable platform for the production of bio-based chemicals and polymers. *Polymer Chem.* **6**: 4497-4559.
- Ismail, I., Hamayun, M., Sayyed, A., Din, I.U., Gul, H., Hussain, A. 2016. Gibberellin and indole acetic acid production capacity of endophytic fungi isolated from *Zea mays* L. *Inter. J. Biosci.* **8**: 35-43.
- Jena, S.K., Tayung, K. 2013. Endophytic fungal communities associated with two ethno-medicinal plants of Similipal Biosphere Reserve, India and their antimicrobial prospective. *J. Appl. Pharmaceut. Sci.* **3**: S7.
- Kaul, S., Ahmed, M., Sharma, T., Dhar, M.K. 2014. Unlocking the myriad benefits of endophytes: an overview. In *Microbial diversity and biotechnology in food security* (R.N. Kharwar, R.S. Upadhyay, N.K. Dubey, R. Raghuvanshieds.), Springer publication, Pp 41-57.
- Kellogg, J.J., Raja, H.A. 2017. Endolichenic fungi: a new source of rich bioactive secondary metabolites on the horizon. *Phytochem. Rev.* **16**: 271-293.
- Khan, A. R., Ullah, I., Waqas, M., Shahzad, R., Hong, S. J., Park, G. S., Shin, J. H. 2015. Plant growth-promoting potential of endophytic fungi isolated from *Solanum nigrum* leaves. *World J. Microbiol. Biotechnol.* **31**: 1461-1466.
- Khan, S.A., Hamayun, M., Yoon, H., Kim, H.Y., Suh, S.J., Hwang, S.K., Kim, J.G. 2008. Plant growth promotion and *Penicillium citrinum*. *BMC Microbiol.* **8**: 1-10.
- KouipouToghueo, R. M., Boyom, F. F. 2019. Endophytic fungi from *Terminalia* species: a comprehensive review. *J. Fungi* **5**: 43.
- Li, Y.L., Xin, X.M., Chang, Z.Y., Shi, R.J., Miao, Z.M., Ding, J., Hao, G.P. 2015. The endophytic fungi of *Salvia miltiorrhiza* are a potential source of natural antioxidants. *Bot. Studies* **56**: 1-7.
- Liu, X., Dong, M., Chen, X., Jiang, M., Lv, X., Zhou, J. 2008. Antimicrobial activity of an endophytic *Xylaria* sp. YX-28 and identification of its antimicrobial compound 7-amino-4-methylcoumarin. *Appl. Microbiol. Biotechnol.* **78**: 241-247.
- Logesh, A.R., Thillaimaharani, K.A., Sharmila, K., Kalaiselvam, M., Raffi, S.M. 2012. Production of chitosan from endolichenic fungi isolated from mangrove environment and its antagonistic activity. *Asian Pac. J. Trop. Biomed.* **2**: 140-143.
- Maehara, S., Simanjuntak, P., Maetani, Y., Kitamura, C., Ohashi, K., Shibuya, H. 2013. Ability of endophytic filamentous fungi associated with *Cinchona ledgeriana* to produce Cinchona alkaloids. *J. Nat. Med.* **67**: 421-423.
- Maehara, S., Simanjuntak, P., Ohashi, K., Shibuya, H. 2010. Composition of endophytic fungi living in *Cinchona ledgeriana* (Rubiaceae). *J. Nat. Med.* **64**: 227-230.
- Mohanta, J., Tayung, K., Mohapatra, U.B. 2008. Antimicrobial potentials of endophytic fungi inhabiting three ethno-medicinal plants of Similipal Biosphere Reserve, India. *Internet J. Microbiol.* **5**: 1-8
- Nath, A., Chattopadhyay, A., Joshi, S. R. 2015. Biological activity of endophytic fungi of *Rauwolfia serpentina* Benth: an ethnomedicinal plant used in folk medicines in Northeast India. *Proc. National Acad. Sci., India Section B: Biol. Sci.* **85**: 233-240.
- Nithya, K., Muthumary, J. 2011. Bioactive metabolite produced by *Phomopsis* sp., an endophytic fungus in *Allamanda cathartica* Linn. *Recent Res. Sci. Technol.* **3**: 44-48.
- Padhi, S., Tayung, K. 2013. Antimicrobial activity and molecular characterization of an endophytic fungus, *Quambalaria* sp. isolated from *Ipomoea carnea*. *Annal. Microbiol.* **63**: 793-800.
- Padhi, S., Tayung, K. 2015. In vitro antimicrobial potentials of endolichenic fungi isolated from thalli of *Parmelia* lichen against some human pathogens. *Beni-Suef Univ. J. Basic Appl. Sci.* **4**: 299-306.
- Padhi, S., Masi, M., Cimmino, A., Tuzi, A., Jena, S., Tayung, K., Evidente, A. 2019. Funiculosone, a substituted dihydroxanthene-1, 9-dione with two of its analogues produced by an endolichenic fungus *Talaromyces funiculosus* and their antimicrobial activity. *Phytochemistry* **157**: 175-183.
- Padhi, S., Masi, M., Panda, S.K., Luyten, W., Cimmino, A., Tayung, K., Evidente, A. 2020. Antimicrobial secondary metabolites of an endolichenic *Aspergillus niger* isolated from lichen thallus of *Parmotrema ravum*. *Nat. Prod. Res.* **34**: 2573-2580.
- Palem, P. P., Kuriakose, G. C., Jayabaskaran, C. 2015. An endophytic fungus, *Talaromyces radicus*, isolated from *Catharanthus roseus*, produces vincristine and vinblastine, which induce apoptotic cell death. *PLoS one* **10**: e0144476.
- Pan, F., Su, X., Hu, B., Yang, N., Chen, Q., Wu, W. 2015. *Fusarium redolens* 6WBY3, an endophytic fungus isolated from *Fritillaria unibracteata* var. *wabuensis*, produces peimisine and imperialine-3- β -D-glucoside. *Fitoterapia* **103**: 213-221.
- Paranagama, P.A., Wijeratne, E.K., Burns, A.M., Marron, M.T., Gunatilaka, M.K., Arnold, A.E., Gunatilaka, A. L. 2007. Heptaketides from *Corynespora* sp. inhabiting the cavern beard lichen, *Usnea cavernosa*: first report of metabolites of an endolichenic fungus. *Journal of natural products* **70**: 1700-1705.
- Photita, W., Lumyong, S., Lumyong, P., Hyde, K. D. 2001. Endophytic fungi of wild banana (*Musa acuminata*) at doi Suthep Pui National Park, Thailand. *Mycol. Res.* **105**: 1508-1513.
- Potshangbam, M., Devi, S.I., Sahoo, D., Strobel, G.A. 2017. Functional characterization of endophytic fungal community associated with *Oryza sativa* L. and *Zea mays* L. *Front. Microbiol.* **8**: 325.
- Rakotoniriana, E. F., Munaut, F., Decock, C., Randriamampionona, D., Andriamboloniaina, M., Rakotomalala, T., Corbisier, A. M. 2008. Endophytic fungi from leaves of *Centella asiatica*: occurrence and potential interactions within leaves. *Antonie van Leeuwenhoek* **93**: 27-36.
- Rana, K.L., Kour, D., Kaur, T., Devi, R., Yadav, A.N., Yadav, N., Saxena, A.K. 2020. Endophytic microbes: biodiversity, plant growth-promoting mechanisms and potential applications for agricultural sustainability. *Antonie Van Leeuwenhoek* **113**: 1075-1107.
- Raviraja, N. S., Maria, G. L., Sridhar, K. R. 2006. Antimicrobial evaluation of endophytic fungi inhabiting medicinal plants of the Western Ghats of India. *Eng. Life Sci.* **6**: 515-520.
- Rodriguez, R.J., White, Jr. J.F., Arnold, A.E., Redman, R.S. 2009. Fungal endophytes: diversity and functional roles. *New Phytol.* **182**: 314-330.
- Roy, S., Mili, C., Talukdar, R., Wary, S., Tayung, K. 2021. Seed Borne Endophytic Fungi Associated with Some Indigenous Rice Varieties of North East India and Their Growth Promotion and Antifungal Potential. *Ind. J. Agric. Res.* **55**: 603-608.
- Seetharaman, P., Gnanasekar, S., Chandrasekaran, R., Chandrakasan, G., Kadarkarai, M., Sivaperumal, S. 2017. Isolation and characterization of anticancer flavone chrysin (5, 7-dihydroxy flavone)-producing endophytic fungi from *Passiflora incarnata* L. leaves. *Ann. Microbiol.* **67**: 321-331.
- Sette, L. D., Passarini, M. R. Z., Delarmelina, C., Salati, F., Duarte, M. C. T. 2006. Molecular characterization and antimicrobial activity of endophytic fungi from coffee plants. *World J. Microbiol. Biotechnol.* **22**: 1185-1195.
- Silva, G.H., de Oliveira, C.M., Teles, H.L., Pauletti, P.M., Castro-Gamboa, I., Silva, D.H., Araujo, A.R. 2010. Sesquiterpenes from *Xylaria* sp., an endophytic fungus associated with *Piper aduncum* (Piperaceae). *Phytochem. Lett.* **3**: 164-167.
- Strobel, G., Daisy, B. 2003. Bioprospecting for microbial endophytes and their natural products. *Microbiol. Mol. Biol. Rev.* **67**: 491-502.
- Sturz, A.V., Nowak, J. 2000. Endophytic communities of rhizobacteria and the strategies required to create yield enhancing associations with crops. *Appl. Soil Ecol.* **15**: 183-190.
- Suryanarayanan, T.S., Rajagopal, K. 2000. Fungal endophytes (Phellomyces) of some tropical forest trees. *Ind. Forester* **126**: 165-170.
- Suryanarayanan, T.S., Thirunavukkarasu, N. 2017. Endolichenic fungi: the lesser known fungal associates of lichens. *Mycology* **8**: 189-196.

- Talukdar, R., Tayung, K. 2021. Endophytic fungal assemblages of *Zanthoxylum oxyphyllum* Edgew. and their antimicrobial potential. *Plant Sci. Today* **8**: 132-139.
- Talukdar, R., Padhi, S., Rai, A.K., Masi, M., Evidente, A., Jha, D.K., Tayung, K. 2021. Isolation and characterization of an endophytic fungus *Colletotrichum coccodes* producing Tyrosol from *Houttuynia cordata* Thunb. using ITS2 RNA secondary structure and molecular docking study. *Front. Bioengin. Biotechnol.* **9**: 484.
- Talukdar, R., Wary, S., Mili, C., Roy, S., Tayung, K. 2020. Antimicrobial secondary metabolites obtained from endophytic fungi inhabiting healthy leaf tissues of *Houttuynia cordata* Thunb., an ethnomedicinal plant of Northeast India. *J. Appl. Pharm. Sci.* **10**: 99-106.
- Tayung, K., Jha, D.K. 2010. Antimicrobial endophytic fungal assemblages inhabiting bark of *Taxus baccata* L. of Indo-Burma mega biodiversity hotspot. *Ind. J. Microbiol.* **50**: 74-81.
- Tayung, K., Barik, B.P., Jha, D.K., Deka, D.C. 2011. Identification and characterization of antimicrobial metabolite from an endophytic fungus, *Fusarium solani* isolated from bark of Himalayan yew. *Mycosphere* **2**: 203-213.
- Tayung, K., Sarkar, M., Baruah, P. 2012. Endophytic fungi occurring in *Ipomoea carnea* tissues and their antimicrobial potentials. *Brazil. Arch. Biol. Technol.* **55**: 653-660.
- Tejesvi, M. V., Mahesh, B., Nalini, M. S., Prakash, H.S., Kini, K. R., Subbiah, V., Shetty, H. S. 2006. Fungal endophyte assemblages from ethnopharmacologically important medicinal trees. *Can. J. Microbiol.* **52**: 427-435.
- Toghueo, R.M.K., Ejiya, I.E., Sahal, D., Yazdani, S. S., Boyom, F.F. 2017. Production of cellulolytic enzymes by endophytic fungi isolated from Cameroonian medicinal plants. *Int. J. Curr. Microbiol. App. Sci.* **6**: 1264-1271.
- Tripathi, M., Joshi, Y. 2015. Endolichenic fungi in Kumaun Himalaya: a case study. In: *Recent Advances in Lichenology* (D.K. Upreti, P. K. Divakar, V. Shukla, R. Bajpai Eds) Springer, New Delhi. Pp. 111-120.
- Tsavkelova, E.A., Cherdyntseva, T.A., Botina, S.G., Netrusov, A.I. 2007. Bacteria associated with orchid roots and microbial production of auxin. *Microbiolog. Res.* **162**: 69-76.
- Wang, F.W., Jiao, R.H., Cheng, A.B., Tan, S.H., Song, Y.C. 2007. Antimicrobial potentials of endophytic fungi residing in *Quercus variabilis* and brefeldin A obtained from *Cladosporium* sp. *World J. Microbiol. Biotechnol.* **23**: 79-83.
- Wang, L.W., Xu, B.G., Wang, J.Y., Su, Z.Z., Lin, F.C., Zhang, C.L., Kubicek, C. P. 2012. Bioactive metabolites from *Phoma* species, an endophytic fungus from the Chinese medicinal plant *Arisaema erubescens*. *Appl. Microbiol. Biotechnol.* **93**: 1231-1239.
- Wang, X. J., Min, C. L., Ge, M., Zuo, R. H. 2014. An endophytic sanguinarine-producing fungus from *Macleaya cordata*, *Fusarium proliferatum* BLH51. *Curr. Microbiol.* **68**: 336-341.
- Waqas, M., Khan, A.L., Kamran, M., Hamayun, M., Kang, S.M., Kim, Y.H., Lee, I.J. 2012. Endophytic fungi produce gibberellins and indoleacetic acid and promotes host-plant growth during stress. *Molecules* **17**: 10754-10773.
- Wu, W., Dai, H., Bao, L., Ren, B., Lu, J., Luo, Y., Liu, H. 2011. Isolation and structural elucidation of proline-containing cyclopentapeptides from an endolichenic *Xylaria* sp. *Journal of natural products* **74**: 1303-1308.
- Wu, Y., Girmay, S., da Silva, V.M., Perry, B., Hu, X., Tan, G. T. 2015. The role of endophytic fungi in the anticancer activity of *Morinda citrifolia* Linn. (Noni). Evidence-Based *Compl. Alter. Med.* **2015**: Article ID 393960, 8 pages.
- Yang, B., Wang, X., Ma, H., Yang, T., Jia, Y., Zhou, J., Dai, C. 2015. Fungal endophyte *Phomopsis liquidambari* affects nitrogen transformation processes and related microorganisms in the rice rhizosphere. *Front. Microbiol.* **6**: 982
- Yang, H., Tan, N., Wu, F., Liu, H., Sun, M., She, Z., Lin, Y. 2012. Biosorption of uranium (VI) by a mangrove endophytic fungus *Fusarium* sp. ZZF51 from the South China Sea. *J. RadioAnalytic. Nucl. Chem.* **292**: 1011-1016.
- Zhang, H.W., Song, Y.C. and Tan, R.X. 2006. Biology and chemistry of endophytes. *Nat. Prod. Rep.* **23**: 753-771.
- Zhao, J., Fu, Y., Luo, M., Zu, Y., Wang, W., Zhao, C., Gu, C. 2012. Endophytic fungi from pigeon pea [*Cajanus cajan* (L.) Millsp.] produce antioxidant cajanin stilbene acid. *J. Agric. Food Chem.* **60**: 4314-4319.